

Non-Nicotinic Therapies for Smoking Cessation

Eric C.K. Siu and Rachel F. Tyndale

Center for Addiction & Mental Health and Department of Pharmacology, University of Toronto, Toronto, Canada; email: eric.siu@utoronto.ca, r.tyndale@utoronto.ca

Annu. Rev. Pharmacol. Toxicol. 2007. 47:541–64

The *Annual Review of Pharmacology and Toxicology* is online at <http://pharmtox.annualreviews.org>

This article's doi:
10.1146/annurev.pharmtox.47.120505.105354

Copyright © 2007 by Annual Reviews.
All rights reserved

0362-1642/07/0210-0541\$20.00

Key Words

bupropion, varenicline, vaccines, rimonabant, CYP2A6, nicotine metabolism

Abstract

Cigarette smoking is the primary cause of numerous preventable diseases; as such, the goals of smoking cessation are both to reduce health risks and to improve the quality of life. Currently, the first-line smoking cessation therapies include nicotine replacement products and bupropion. The nicotinic receptor partial agonist varenicline has recently been approved by the FDA for smoking cessation. A newer product currently under development and seeking approval by the FDA are nicotine vaccines. Clonidine and nortriptyline have demonstrated some efficacy but side effects may limit their use to second-line therapeutic products. Other therapeutic drugs that are under development include rimonabant, mecamylamine, monoamine oxidase inhibitors, and dopamine receptor D3 antagonists. Inhibitors of nicotine metabolism are also promising candidates for smoking reduction and cessation. In conclusion, promising new therapeutic products are emerging and they will provide smokers additional options to assist in achieving smoking cessation.

INTRODUCTION

It was more than four decades ago when the first Surgeon General's Report on Smoking and Health provided irrefutable evidence that smoking is a cause of many cancers and other diseases (http://www.cdc.gov/Tobacco/sgr/sgr_1964/sgr64.htm). Forty years later, tobacco smoking remains the single most preventable cause of death in the world, responsible for the death of one in ten adults annually (<http://www.who.int/tobacco/en/>). Despite successes in the public health and legislative forefronts [WHO Framework Convention on Tobacco Control (<http://www.fctc.org>)], hundreds of millions of smokers suffer from the consequences of cigarette smoking. Studies have suggested that the cumulative risks of death owing to cardiovascular and lung diseases can be drastically reduced if smokers stop smoking, even late in life (1, 2). Therefore, smoking cessation and harm reduction are means to reduce health risks associated with smoking and to improve the quality of life.

Smoking is a complex and regulated behavior that is influenced by both genetic and environmental factors (3, 4). Nicotine is the primary component of tobacco that is responsible for reward and reinforcement, as well as the withdrawal effects of cigarettes (3, 5, 6). Nicotine activates nicotinic acetylcholine receptors (nAChRs) in the CNS, and one physiological effect of nAChR activation is dopamine release in the nucleus accumbens, a region important for the rewarding properties of the drug (7). Tobacco dependence is due, in part, to both acute and chronic tolerance to nicotine effects; acute tolerance to nicotine is associated with activation and desensitization (8–10) of nAChRs, whereas chronic tolerance is related to receptor upregulation (11–16). Due to the central role of nicotine on reward pathway activation, the degree to which a smoker achieves “satisfaction” from smoking is to a large extent related to the amount of nicotine obtained from the cigarette (6). Smokers titrate their smoking behaviors to maintain an optimal amount of nicotine in the plasma and brain (5, 17, 18); therefore, the number of cigarettes smoked (nicotine acquired) is influenced by the systemic availability of nicotine (19). These findings provided the rationale for the use of nicotine as a means to reduce smoking.

Nicotine replacement products are the first line treatments for nicotine dependence approved by the U.S. Federal Drug Administration (FDA). Currently, there are many nicotine delivery devices available on the market—these include gum, transdermal patch, vapor inhaler, nasal spray, lozenge, and sublingual tablet. These treatments enhance cessation by delivering nicotine without the exposure to carcinogenic compounds found in cigarette smoke. The use of these nicotine replacement products has led to varying degrees of success in long-term smoking cessation (20, 21) which has been reviewed extensively elsewhere (22–24). The only non-nicotine medications approved by the FDA for smoking cessation are the antidepressant bupropion (Zyban, Wellbutrin) and the nicotinic receptor partial antagonist varenicline (Chantix). In addition to these therapies, other pharmacological compounds are also being developed as adjuncts to smoking cessation. Newer treatments in development include nicotine vaccines and rimonabant. At the time of preparation of this review, one of the nicotine vaccines (Nabi Biopharmaceuticals) had received fast-track review status by the FDA.

BUPROPION

Smoking and depression are highly associated, with both being influenced by dopamine levels (25, 26). As a result, a number of antidepressants have been examined for their effects on cigarette consumption. The atypical antidepressant bupropion is the only non-nicotine product approved as a first-line therapy for smoking cessation. The precise mechanisms of bupropion that are responsible for smoking cessation are not well understood; however, the effect may be partly related to the bupropion's ability to reduce cravings in abstinent smokers and to alleviate certain symptoms associated with withdrawal (27–30). Bupropion, at therapeutic doses, can bind to striatal dopamine transporters (~20% occupancy) and potentially prevent dopamine reuptake (31, 32), thereby reducing negative withdrawal symptoms. The noradrenergic system is also implicated in the pharmacological effects of bupropion. At pharmacologically relevant concentrations bupropion reduces the firing rates of noradrenergic neurons in the locus coeruleus (33), which contains neuronal projections to the hippocampus, a region implicated in drug dependence (34). However, the precise mechanism of bupropion's effects on the function of the noradrenergic system is unclear. Bupropion can also act as a noncompetitive antagonist of nicotinic receptors, suggesting that bupropion may attenuate the rewarding effect of nicotine (35–37). Finally, the metabolites of bupropion, in particular hydroxybupropion, have been demonstrated to be pharmacologically active (38); the individual effects of these metabolites on nicotine dependence have not been clarified (38, 39). The vast majority of studies have demonstrated that bupropion is more effective at improving smoking cessation than placebo (27, 40–48), with some studies showing no significant improvement in cessation rates (49, 50). Over the past few years a large amount of data has been gathered with regards to the safety of bupropion use for smoking cessation in a community setting. Overall, the drug caused few serious adverse reactions—the primary reasons associated with discontinuation of treatments were insomnia, nausea/vomiting, and dizziness (51).

VARENICLINE

Varenicline is a partial agonist of the $\alpha 4\beta 2$ nicotinic cholinergic receptor; the structure of varenicline is based on the alkaloid extract cytisine from the plant *Cytisus laburnum* L. (52). As with varenicline, cytisine is also a partial agonist of the $\alpha 4\beta 2$ receptor (53). In parts of Europe, extracts of this plant (Tabex) are used as smoking cessation treatment (54, 55). In mouse studies, the $\beta 2$ receptor subunit is required for nicotine reward as measured by conditioned place preference and receptor upregulation during chronic nicotine treatment, and the $\alpha 4$ receptor subunit is important for reward and development of tolerance to nicotine (56–58). Varenicline was hypothesized to reduce nicotine-mediated activation of the dopaminergic system via partial blockade of receptor stimulation by high levels of nicotine during smoking, thus lowering the rewarding effects of nicotine. In addition, by acting as a weak agonist, varenicline could also stimulate dopamine release in the mesolimbic region and reduce craving and withdrawal during abstinence (52, 53).

In a Phase II study, it was found that the carbon monoxide-confirmed 4-week continuous quit rates between weeks 9 to 12 were significantly higher in smokers treated with varenicline at 0.5 mg b.i.d. (45.1%) and at 1 mg b.i.d. (50.6%) compared with placebo (12.4%) (59). In another Phase II study, varenicline (1 mg b.i.d.) was found to be superior to bupropion (150 mg b.i.d.) in confirmed 4-week continuous quit rates (40.8% versus 28.6%) (60). At week 7, the carbon monoxide-confirmed 7-day point prevalence abstinence was higher for varenicline (1 mg b.i.d.) compared with bupropion (150 mg b.i.d.), and both showed statistically greater response rates than placebo (~52% and ~38% versus ~23%) (60). Subsequently, in one Phase III study, patients were given either varenicline (1 mg b.i.d.), bupropion (150 mg b.i.d.), or placebo. The varenicline group had ~1.5- and ~2.4-times greater continuous abstinence between weeks 9 to 12 compared with the bupropion and placebo groups, respectively (61). When examining the total abstinence between weeks 9 to 52, the varenicline group showed ~1.5- and ~2.5-times greater abstinence compared with the bupropion and placebo groups, respectively (61). However, another Phase III study with identical design did not find a significant difference between varenicline and placebo (61). In another study, maintenance of abstinence was compared between the patients that received 12 weeks of varenicline and patients that received an additional 12 weeks (24 weeks total) of treatment. Results indicated that 70% of the patients receiving 24 weeks of treatment were abstinent between weeks 13 to 24, whereas only 50% of the 12-week-treatment patients were abstinent (61). When examining the subjective effects associated with smoking, results obtained from the Phase III studies described above showed that varenicline provided significantly greater benefit compared with placebo. Areas of benefit, using the modified cigarette evaluation questionnaire [adopted from Rose et al. (102)], included psychological reward, smoking satisfaction, enjoyment of respiratory tract sensation, and craving reduction, but not aversion (62). In contrast, bupropion only showed benefit in the psychological reward subscale compared with placebo (62).

The safety of varenicline (compared with bupropion and placebo) has also been evaluated (60). Overall, the adverse events observed, or reasons for discontinuing treatment, were relatively mild. The percentage of patients who discontinued the study owing to adverse events was similar between varenicline (1 mg b.i.d.; 11.2%) and bupropion (150 mg b.i.d.; ~15.9%) compared with placebo (~9.8%). The most common adverse event associated with varenicline treatment was nausea (52.0% versus 21.4% and 18.7% for bupropion and placebo, respectively) (60). In a long-term (52-week) study, the primary adverse events contributing to patient dropout were nausea, abnormal dreams, and insomnia (7.6%, 2.4%, and 3.2% for varenicline, bupropion, and placebo, respectively) (60). The overall discontinuation rates were significantly higher at 28.3% in the varenicline group compared with 10.3% in the placebo group (63). The results from Phase III studies indicated that varenicline can provide greater smoking cessation compared with bupropion; however, safety issues associated with long-term use of this drug remain to be determined if it is to be used for longer than 24 weeks.

NICOTINE VACCINES

A newer strategy currently being investigated for smoking cessation is the nicotine vaccine. The principle of this strategy is to prevent nicotine from entering the brain. In immunized individuals, nicotine obtained from smoking is bound by nicotine-specific antibodies and cannot cross the blood-brain barrier, thus preventing its central effects (64, 65). Nicotine alone is not immunogenic (i.e., it does not elicit antinicotine antibody production); therefore, it must be conjugated to larger carrier proteins that can act as immunogenic molecules. Currently, there are at least five companies developing nicotine vaccines (Cytos Biotechnology, Nabi Biopharmaceuticals, Xenova Group Ltd., Chilka Ltd., and Independent Pharmaceutica AB). Each company uses a specific antigenic molecular approach. For example, Cytos utilizes a virus-like particle as the immunogen, whereas Nabi selected recombinant exoprotein A and Xenova makes use of a recombinant cholera toxin B (66).

An advantage of nicotine vaccines, compared with other forms of pharmacotherapy, is that daily administration of the drug is not required; only occasional booster shots are needed to maintain an adequate antibody titer. However, there are also potential drawbacks to nicotine vaccines and these have been discussed extensively elsewhere (67, 68). One of the primary concerns associated with the nicotine vaccine is that the titer of antibodies after immunization may not be sufficient to sequester all of the nicotine in arterial blood, limiting its use to prevent nicotine entry into the brain during smoking (64, 65, 67). If small amounts of nicotine enter the brain with each cigarette, the vaccine may in fact increase smoking to compensate for reduced nicotine levels (67, 68).

In animal studies, immunization with nicotine vaccines prevented the reinstatement of nicotine self-administration in rats when reexposed to nicotine after extinction of the behavior (69). However, another study found that vaccination did not prevent acquisition of nicotine self-administration, but reduced self-administration once self-administration was acquired (70). One peer-reviewed study reported the effects of vaccination on smoking behaviors (NicVAX by Nabi Biopharmaceutical) (71). In this 38-week study with ad libitum smoking it was found that the nicotine vaccine, at all doses tested (50, 100, and 200 μ g; 1 primary challenge shot and 3 booster shots at each dose), did not cause compensatory smoking behaviors or precipitate withdrawal, the latter being consistent with animal data (72). In addition, the 30-day continuous abstinence rates were higher in patients receiving the highest vaccine dose (~38%) compared with placebo (~10%) (71). Careful interpretation of these results is needed, as the sample size was very small and the study was designed to evaluate the safety and immunogenicity of the vaccine rather than smoking cessation efficacy.

Cytos Biotechnology recently posted findings from a Phase II study regarding the efficacy of their vaccine on smoking cessation on the company's Web site (<http://www.cytos.com>; accessed on April 19, 2006). They reported that a larger portion of high responders (high-titer production) were continuously abstinent from smoking compared with the placebo group at 6 months (57% versus 31%) and at 12 months (42% versus 21%). These studies show promise and suggest that the vaccine may be useful for smoking cessation treatment (and possibly for relapse prevention).

Thus far, there have been no serious adverse events associated with the vaccines. The majority (>99%) of the reported side effects were mild to moderate, with injection-site reaction, flu-like symptoms, headache, and myalgia being the most common. All effects were resolved without medical treatments (71, 73).

A particular concern associated with active immunization is whether the resulting antibody titer is sufficient to prevent all nicotine from entering the brain; rats treated with the vaccine only showed 64% reduction in brain nicotine (from 2 cigarettes equivalent of nicotine) (65). One way to circumvent the problem associated with antibody titer is through passive immunization, in which nicotine-specific antibodies are administered directly. A recent study has shown that immunization of rats with high doses (240 mg/kg) of a low-affinity monoclonal, or a high-affinity polyclonal, antibody significantly reduced nicotine distribution into the brain (up to ~75% reduction with the low-affinity monoclonal antibody) (74). This effect appeared to be dose related, with no significant difference between high- and low-affinity antibodies (74). Functionally, passive immunization prevented nicotine-conditioned place preference, reduced the development of nicotine dependence, and prevented the ability of nicotine to reduce abstinence symptoms (70, 75, 76). However, its effects on nicotine self-administration in animals, and smoking in humans, are not yet known. Nicotine antibodies may provide a potential alternative to active vaccines; however, the high costs associated with antibody production may be prohibitive.

RIMONABANT (SR141716)

The possible involvement of the endocannabinoid system in drug dependence was suggested when it was found that delta 9-tetrahydrocannabinol and related compounds could trigger dopamine efflux in the mesolimbic system (77–79). Later, the endocannabinoid system was implicated in nicotine dependence when the levels of endogenous cannabinoid-like compounds were found to be altered during chronic nicotine administration (80). Specifically, nicotine increased levels of endogenous agonists of the CB₁ and CB₂ receptors [anandamide (AEA) and 2-arachidonol-glycerol (2-AG), respectively] in the brainstem, a region involved in drug withdrawal (80–82). AEA levels were also increased in the limbic forebrain; however, both AEA and 2-AG levels were decreased in the hippocampus and cerebral cortex. No changes in the CB₁ cannabinoid receptor levels were seen in these brain regions (80). More definitive experiments demonstrating the role of endocannabinoid receptors in nicotine reward were conducted in rats. The CB₁ receptor antagonist SR141716 (rimonabant) prevented IV nicotine self-administration, blocked the release of dopamine in the nucleus accumbens in response to nicotine (83), and prevented both conditioned stimuli-induced nicotine-seeking behaviors and nicotine-conditioned place preference (84, 85). Finally, CB₁ receptor knockout mice showed nicotine-conditioned place preference in one study but not another, which may have been due to differences in dose and the route of administration (86, 87). The precise mechanism of actions of CB₁ receptor antagonism on nicotine reward/reinforcement remains unclear.

In a Phase III study (STRATUS-US) examining the effect of rimonabant on smoking cessation, smokers were given placebo, 5 mg rimonabant, or 20 mg rimonabant

once daily (88). Patients were treated for 10 weeks total, 2 weeks prior to the quit date and 8 weeks following the quit date. Smoking cessation was evaluated during the last four weeks of treatment. It was found that significantly more (27.6%) patients receiving 20 mg rimonabant remained abstinent compared with those who were on 5 mg (15.6%) or placebo (16.1%) (88). In contrast, another study (STRATUS-Europe) found no significant increase in cessation for either treatment dose groups (5 and 20 mg) compared with the placebo group (24%, 25%, and 20%, respectively) (89). In a third and larger study (STRATUS-Worldwide) examining smoking relapse, significantly more patients who received 20 mg rimonabant (~42%) remained smoke-free for the entire year (89) compared with patients receiving placebo treatment (~32%). Rimonabant was well tolerated with discontinuation from the study (STRATUS-US) due to adverse events occurring in 6.9%, 6.1%, and 4.2% of patients receiving 20 mg, 5 mg, and placebo, respectively. The most common side effects were nausea and upper respiratory tract infection (88). Rimonabant has not yet been approved for use as a smoking cessation product in the United States and the moderate benefits obtained from this drug may limit its widespread use.

OTHER NON-NICOTINIC PHARMACOTHERAPIES

Clonidine and nortriptyline are second-line therapies for smoking cessation that are being tested but have not yet been approved for this indication by the FDA. The noradrenergic tricyclic antidepressant nortriptyline inhibits serotonin and noradrenalin reuptake. It has demonstrated some efficacy in smoking cessation (90, 91), likely by reducing withdrawal symptoms. Odds ratios for smoking cessation using nortriptyline alone in four studies ranged from 1.2 to 5.5, with only one study lacking a statistical significant benefit (90–93). The smoking cessation rates achieved with nortriptyline appear to be comparable to those achieved with bupropion [pooled odds ratios: 2.1 (1.5–3.1) versus 2.0 (1.7–3.4)] (94). Nonetheless, the current lack of extensive testing and safety information for smoking cessation limits the use of nortriptyline as second-line therapy (94).

Clonidine is an α -2-adrenergic receptor agonist used primarily for the treatment of hypertension, although it has also been used for treatment of opiate and alcohol withdrawal (95, 96). Clonidine may alleviate withdrawal symptoms associated with tobacco use (97, 98). A recent Cochrane analysis reviewed the efficacy of clonidine for smoking cessation from six studies and found that clonidine treatment was associated with increased smoking cessation [pooled odds ratio of 1.89 (95% CI: 1.30–2.74)]; however, when analyzed separately only one study showed significant improvement in smoking cessation (99). Furthermore, adverse effects such as dry mouth, sedation, and postural hypotension limited its general use.

Mecamylamine is a nicotinic antagonist originally used as an antihypertensive (100). At low doses, mecamylamine is effective at blocking the rewarding effects of nicotine and enhancing short-term smoking cessation, especially in conjunction with transdermal nicotine (101, 102). Moreover, although an antagonist of nicotinic receptors, low doses of oral mecamylamine did not appear to precipitate withdrawal in

smokers (103). Additional studies are required to ascertain the long-term effectiveness of mecamylamine in smoking cessation.

Other compounds that have been assessed for smoking cessation include the monoamine oxidase inhibitors (MAOIs). Cigarette smoking has been demonstrated to inhibit MAO-A and MAO-B, potentially by reducing metabolism of dopamine (104). A preliminary study found that the reversible MAO-A inhibitor moclobemide significantly reduced self-reported smoking rates, although several individuals who reported abstinence had higher than acceptable plasma levels of the nicotine metabolite cotinine, suggesting continued smoking (105). The irreversible MAO-B inhibitor selegiline has also been demonstrated to provide some relief from smoking, but this effect was short term with no significant benefit in the treatment group compared with the placebo group in the last four weeks of an eight-week study (106). The reversible inhibitor of MAO-B lazabemide was also evaluated for smoking cessation, although the study was terminated due to concerns of hepatotoxicity (107).

Emerging evidence indicates that the dopamine receptor D3 (DRD3) is involved in the motivational aspects of drug-seeking behaviors (108–112). One study showed that the DRD3-selective antagonist SB-277011-A reduced the relapse to nicotine self-administration triggered by nicotine in rats but it did not affect nicotine self-administration (108). Another study found that the DRD3 partial agonist BP-897 blocked nicotine-conditioned place preference in rats (111). Furthermore, DRD3 appears to be involved in conditioned stimulus associated with addictive substances but not natural reinforcers (e.g., food and sex) (113). Therefore, modulators of DRD3 may be useful for relapse prevention in smoking cessation (114).

INHIBITORS OF NICOTINE METABOLISM

Nicotine Metabolism

Because nicotine elimination is a key factor in determining the number of cigarettes smoked, a novel strategy that may be useful for smoking cessation is to reduce the elimination of nicotine, and one way to achieve this is to inhibit its metabolism. In humans, approximately 80% of nicotine is metabolized to cotinine (115). Approximately 90% of the conversion of nicotine to cotinine is mediated by the hepatic enzyme CYP2A6 (116). This conversion is a two-step process that involves the initial formation of nicotine- $\Delta^{-1(5)}$ iminium ion by CYP2A6 (117, 118). The iminium ion is then converted into cotinine by the cytosolic enzyme aldehyde oxidase (116, 119). Nicotine is also excreted unchanged and metabolized to other minor metabolites, but these pathways account for only a small portion of the metabolic fate of nicotine (120–124). Cotinine is further metabolized to *trans*-3-hydroxycotinine, mediated solely by CYP2A6 (125, 126). As CYP2A6 is the primary enzyme that metabolizes nicotine, alterations (e.g., inhibition) in the amount or function of this enzyme will significantly affect plasma nicotine levels during smoking and during NRT treatment and thus may alter smoking behaviors and the efficacy of NRTs (127, 128).

Genetic Variation in *CYP2A6* and the Impact on Nicotine Metabolism and Smoking Behaviors

CYP2A6 is a highly polymorphic gene; currently, there are 23 numbered *CYP2A6* allelic variants and many additional single nucleotide polymorphisms (<http://www.imm.ki.se/CYPalleles/cyp2a6.htm>). Many of the genetic variants have been shown to alter the expression, stability, and function of the *CYP2A6* enzyme. For instance, *CYP2A6*4* is a gene deletion variant where no enzyme is produced; individuals with two alleles of this variant have impaired nicotine metabolism forming very low levels of cotinine (129–131). In terms of smoking behaviors, slower metabolizers (individuals with $\leq 50\%$ in nicotine metabolism activity) have reduced smoking behaviors, such as consuming fewer cigarettes per day (as indicated by self-report and lower carbon monoxide levels), and are at lower risk for being dependent smokers (128, 132, 133). Slower metabolizers also smoke for shorter durations and were more likely to quit smoking compared with normal metabolizers (133, 134).

Inhibition of *CYP2A6* Activity

Based on the effect of *CYP2A6* genotypes on the nicotine metabolism and smoking behaviors, it was hypothesized that one could reduce smoking by mimicking defects in the *CYP2A6* enzyme (phenocopying). Furthermore, if inhibition of nicotine metabolism can mimic a gene defect, the long-term behavioral effects of these inhibitors may be predicted based on available pharmacogenetic and behavioral studies. *CYP2A6* inhibitors studied in experimental settings (for inhibition of nicotine metabolism and reduction of smoking behaviors) include methoxsalen (8-methoxypsoralen, 8-MOP) and tranylcypromine (TCP). Methoxsalen is both a competitive and a mechanism-based inhibitor of *CYP2A6*, whereas TCP is a competitive inhibitor (135–138). Both drugs are potent and selective inhibitors, although at higher concentrations they can inhibit other CYPs (135, 138–141).

Improving Smoking Cessation Through *CYP2A6* Inhibition

By mimicking defects in nicotine metabolism, one can effectively increase the systemic availability of coadministered nicotine. In practical terms, concurrent use of inhibitors of *CYP2A6* with NRT products should enhance smoking cessation. For example, users of nicotine gum may benefit from the concomitant use of a *CYP2A6* inhibitor. Much of the nicotine released during the chewing of nicotine gum is absorbed by the buccal (oral) cavity, while a significant portion is swallowed (142). Of the nicotine swallowed (as in nicotine gum or in a pill form), approximately 80% is metabolized before entering systemic circulation via first-pass metabolism (143). Thus, inhibition of this first-pass metabolism with a *CYP2A6* inhibitor would increase the nicotine acquired from the gum. In a human study examining the effect of the *CYP2A6* inhibitor methoxsalen on nicotine levels obtained from nicotine gum, it was found that following three days of methoxsalen (10 mg t.i.d.) plus nicotine

gum (4 mg), the mean plasma nicotine levels were significantly higher compared to a placebo inhibitor plus nicotine gum (15.3 versus 10.1 ng/ml; $p < 0.01$) (144).

An oral (pill) form of nicotine is not currently available despite this being a preferred route of drug administration. The large first-pass metabolism prevents pharmacologically relevant levels of nicotine from reaching the systemic system. Higher doses of oral nicotine, to overcome first-pass metabolism, are not possible as higher concentrations of nicotine can cause gastrointestinal irritation (143). Therefore, it may be possible to increase the efficacy of oral nicotine by inhibiting the first-pass metabolism. In a human experimental study, it was found that the addition of the CYP2A6 inhibitor methoxsalen (10 and 30 mg) to 4 mg of oral nicotine significantly increased the subjects' mean plasma nicotine level ($>70\%$) and decreased the subjects' desire to smoke compared with oral nicotine alone ($p < 0.05$) (145). In another study, oral nicotine (4 mg) and methoxsalen (30 mg) significantly reduced the number of cigarettes smoked, level of breath carbon monoxide, number of puffs, as well as inhalation intensity compared with placebo. The subjects also had increased latency to lighting the second cigarette (146).

Administration of TCP (2.5 and 10 mg) increased the plasma concentrations of nicotine by 43% and 65%, respectively, in abstinent smokers receiving 4 mg of oral nicotine and reduced the subjects' desire to smoke (145). These studies provided the first evidence that inhibiting nicotine metabolism could reduce smoking behaviors. Nonetheless, further studies are needed to examine the long-term cessation rates in individuals treated with CYP2A6 inhibitors and oral nicotine.

Although methoxsalen is effective at reducing smoking behaviors in the presence of oral nicotine, CYP2A6 inhibition alone may also reduce smoking and increase quitting, similar to the pharmacogenetic findings in slow metabolizers (128, 132–134); however, this remains to be tested. One company (Nicogen) is currently conducting Phase II clinical trials for smoking reduction and cessation using these approaches.

Tobacco-Specific Nitrosamines

One of the most potent procarcinogens present in tobacco smoke is 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK). The primary metabolic detoxification pathway of NNK is the conversion to 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanol (NNAL), which is then conjugated to NNAL-glucuronide (147, 148). Conversely, NNK can also be activated, via α -hydroxylation, to a reactive metabolite capable of forming DNA adducts (149, 150). CYP2A6 has been demonstrated to be one of the enzymes responsible for this α -hydroxylation (151). Consistent with this, a recent epidemiology study found that individuals that are *CYP2A6*-poor metabolizers have significantly lower risk for developing lung cancer, even after adjusting for the number of cigarettes smoked (152). In animal models, exposure to NNK leads to formation of lung tumors (153–155) and studies in mice have shown that administration of methoxsalen reduced the formation of NNK-induced lung tumors (156, 157). Administration of methoxsalen in human smokers rerouted excretion of NNK as increased NNAL and NNAL-glucuronide, suggesting significant inhibition of the α -hydroxylation of NNK (158). Therefore, in addition to enhancing

smoking cessation, inhibitors of nicotine metabolism can potentially provide harm reduction in smokers attempting to quit.

Discovery of Novel CYP2A6 Inhibitors

Both quantitative structure-activity relationship models for CYP2A6 inhibitors and the crystal structure of CYP2A6 are available (159–161). It has been predicted that favorable substrates for CYP2A6 are small, hydrophilic, and exhibit the ability to adopt a planar structure (159, 160, 162). Over the past few years a large number of compounds have been tested for their ability to inhibit CYP2A6 activity (138, 160, 163–165). The compounds that are most potent and selective inhibitors of CYP2A6 were found to be structural analogues of nicotine and coumarin (160, 165). It is conceivable that novel compounds can be developed that can act as both inhibitors of nicotine metabolism and partial agonists of nicotinic acetylcholine receptors that can attenuate the reinforcing properties of nicotine (53). The combination of CYP2A6 structural information and improvements in QSAR analysis techniques should greatly enhance the ability to discover and develop specific and potent inhibitors of CYP2A6.

Animal Models for Testing Novel CYP2A6 Inhibitors

With an increasing number of potential CYP2A6 inhibitors being discovered (138, 160, 163–165), a more practical model is needed to test their specificity, efficacy, toxicity profile, and impact on nicotine-mediated behaviors. Rodents are the most frequently used animal models for studying the pharmacology of nicotine. While these models are invaluable for understanding the underlying mechanisms of nicotine dependence, there are clear limitations that should be considered. For instance, rats are not a suitable model for studying inhibitors of nicotine metabolism by CYP2A6 because the primary enzymes that metabolize nicotine in these animals belong to the CYP2B family (166, 167). In mice, the majority of nicotine appears to be metabolized by CYP2A5 (118). The mouse CYP2A5 has 86% amino acid sequence similarity to human CYP2A6. From the point of view of drug development, extrapolation of animal data to humans requires caution because interspecies differences can translate to differing effectiveness of the inhibitors on nicotine metabolism. Mice transplanted with human hepatocytes (replacing more than 80% of mouse hepatocytes) have been generated (168–171) and this model may represent an optimal strategy for the evaluation of inhibitors of nicotine metabolism *in vivo* and their effects on nicotine-mediated behaviors.

CONSIDERATION OF NONPHARMACOLOGICAL FACTORS IN SMOKING CESSATION

Although the central pharmacological effects of nicotine are crucial for tobacco dependence, nicotine is not solely responsible for human smoking behaviors. Many studies have examined the contribution of nonpharmacological aspects of cigarette smoking to tobacco dependence (reviewed extensively in Reference 172). For instance,

neither pure nicotine nor denicotinized cigarettes alone completely abolished cravings or provided the subjective satisfaction of smoking regular cigarettes (173, 174). The nonpharmacological components of tobacco dependence have been partly attributed to the sensory aspects of cigarette smoke (e.g., taste, pharyngeal/tracheobronchial sensory cues). Administration of local anesthetic to the airway significantly reduced craving as well as satisfaction from smoking (175, 176) and blockade of olfactory/taste stimuli from smoking reduced self-administration of cigarette puffs, especially in women (177). Together, these findings indicate that non-nicotinic factors play an important role in smoking, which may reduce the success of NRTs in smoking cessation (178).

Despite the accumulating evidence suggesting the importance of nonpharmacological factors in tobacco dependence, there are currently no treatments that specifically target these components. There have been attempts to design non-nicotine inhaler/aerosol products that mimic the sensations of cigarette smoking (179, 180); but thus far no commercial products have been developed. Denicotinized cigarettes are successful at relieving some of the cigarette cravings, therefore they may be useful for dissociating the reinforcing properties of cigarette smoke (sensory cues) from that of nicotine (173, 174). However, the harmful effects (from cigarette smoke) may limit this approach. Finally, behavioral counseling can help patients stop smoking, and this should be combined with newer pharmacological treatments to determine if greater benefits exist compared to either therapy alone (181, 182).

FUTURE PERSPECTIVES

The first Surgeon General's Report on Smoking and Health Tobacco stated that "cigarette smoking is a health hazard of sufficient importance in the United States to warrant appropriate remedial action" (http://www.cdc.gov/Tobacco/sgr/sgr_1964/sgr64.htm). Indeed, the health consequences of tobacco smoking are staggering and truly effective strategies to reduce smoking and to improve public health are needed. Tobacco dependence is a chronic disease of a complex nature but thus far the primary treatments are the NRTs, which deliver only one component responsible for the addictive properties of cigarettes. Although NRTs are important in reducing harm exposure and can improve the overall successful quit rates, many smokers who quit eventually relapse, even with relapse prevention interventions (183). Therefore, to achieve better smoking cessation rates, more comprehensive treatment strategies are required. Antidepressants such as bupropion can reduce cravings as well as withdrawal symptoms. The long-term benefits of the newly approved drug varenicline remain to be seen. Newly developed drugs such as nicotine vaccines are seeking approval for smoking cessation and their long-term benefits compared with existing treatments are not yet determined. Inhibition of nicotine metabolism as a means for smoking cessation shows promising results, but the long-term efficacy remains to be determined. Finally, nonpharmacological factors associated with tobacco dependence need to be addressed as they appear to be crucial for the addictive properties of cigarettes.

DISCLOSURE STATEMENT

R.F.T. is a shareholder in Nicogen, a company focused on novel treatment approaches involving inhibition of hepatic CYP2A6.

LITERATURE CITED

1. Peto R, Darby S, Deo H, Silcocks P, Whitley E, Doll R. 2000. Smoking, smoking cessation, and lung cancer in the UK since 1950: combination of national statistics with two case-control studies. *BMJ* 321:323–29
2. Anthonisen NR, Skeans MA, Wise RA, Manfreda J, Kanner RE, Connett JE. 2005. The effects of a smoking cessation intervention on 14.5-year mortality: a randomized clinical trial. *Ann. Intern. Med.* 142:233–39
3. Henningfield JE, Keenan RM. 1993. Nicotine delivery kinetics and abuse liability. *J. Consult. Clin. Psychol.* 61:743–50
4. Maes HH, Sullivan PF, Bulik CM, Neale MC, Prescott CA, et al. 2004. A twin study of genetic and environmental influences on tobacco initiation, regular tobacco use and nicotine dependence. *Psychol. Med.* 34:1251–61
5. Benowitz NL. 1988. Drug therapy. Pharmacologic aspects of cigarette smoking and nicotine addiction. *N. Engl. J. Med.* 319:1318–30
6. Scherer G. 1999. Smoking behavior and compensation: a review of the literature. *Psychopharmacology* 145:1–20
7. Balfour DJ. 2004. The neurobiology of tobacco dependence: a preclinical perspective on the role of the dopamine projections to the nucleus accumbens [corrected]. *Nicotine Tob. Res.* 6:899–912
8. James JR, Villanueva HF, Johnson JH, Arezo S, Rosecrans JA. 1994. Evidence that nicotine can acutely desensitize central nicotinic acetylcholinergic receptors. *Psychopharmacology* 114:456–62
9. Robinson SE, James JR, Lapp LN, Vann RE, Gross DF, et al. 2006. Evidence of cellular nicotinic receptor desensitization in rats exhibiting nicotine-induced acute tolerance. *Psychopharmacology* 184:306–13
10. Gentry CL, Lukas RJ. 2002. Regulation of nicotinic acetylcholine receptor numbers and function by chronic nicotine exposure. *Curr. Drug Targets CNS Neurol. Disord.* 1:359–85
11. Breese CR, Marks MJ, Logel J, Adams CE, Sullivan B, et al. 1997. Effect of smoking history on [3H]nicotine binding in human postmortem brain. *J. Pharmacol. Exp. Ther.* 282:7–13
12. Marks MJ, Romm E, Gaffney DK, Collins AC. 1986. Nicotine-induced tolerance and receptor changes in four mouse strains. *J. Pharmacol. Exp. Ther.* 237:809–19
13. Marks MJ, Stitzel JA, Collins AC. 1986. Dose-response analysis of nicotine tolerance and receptor changes in two inbred mouse strains. *J. Pharmacol. Exp. Ther.* 239:358–64
14. Perry DC, Davila-Garcia MI, Stockmeier CA, Kellar KJ. 1999. Increased nicotinic receptors in brains from smokers: membrane binding and autoradiography studies. *J. Pharmacol. Exp. Ther.* 289:1545–52

15. Nguyen HN, Rasmussen BA, Perry DC. 2004. Binding and functional activity of nicotinic cholinergic receptors in selected rat brain regions are increased following long-term but not short-term nicotine treatment. *J. Neurochem.* 90:40–49
16. Parker SL, Fu Y, McAllen K, Luo J, McIntosh JM, et al. 2004. Up-regulation of brain nicotinic acetylcholine receptors in the rat during long-term self-administration of nicotine: disproportionate increase of the alpha6 subunit. *Mol. Pharmacol.* 65:611–22
17. Henningfield JE, Goldberg SR. 1988. Pharmacologic determinants of tobacco self-administration by humans. *Pharmacol. Biochem. Behav.* 30:221–26
18. Russell MA, Feyerabend C. 1978. Cigarette smoking: a dependence on high-nicotine boli. *Drug. Metab. Rev.* 8:29–57
19. Le Houezec J. 2003. Role of nicotine pharmacokinetics in nicotine addiction and nicotine replacement therapy: a review. *Int. J. Tuberc. Lung. Dis.* 7:811–19
20. Stead L, Lancaster T. 2005. Nicotine replacement therapy for smoking cessation: Cochrane systematic review. *Int. J. Epidemiol.* 34:1001–2; discussion 2, 3
21. Silagy C, Lancaster T, Stead L, Mant D, Fowler G. 2004. Nicotine replacement therapy for smoking cessation. *Cochrane Database Syst. Rev.* 3:CD000146
22. Frishman WH, Mitta W, Kupersmith A, Ky T. 2006. Nicotine and non-nicotine smoking cessation pharmacotherapies. *Cardiol. Rev.* 14:57–73
23. Henningfield JE, Fant RV, Buchhalter AR, Stitzer ML. 2005. Pharmacotherapy for nicotine dependence. *CA Cancer J. Clin.* 55:281–99; quiz 322–23, 25
24. Mitrouska I, Bouloukaki I, Siafakas NM. 2006. Pharmacological approaches to smoking cessation. *Pulm. Pharmacol. Ther.* In press
25. Paperwalla KN, Levin TT, Weiner J, Saravay SM. 2004. Smoking and depression. *Med. Clin. North Am.* 88:1483–94, x–xi
26. Quattrochi E, Baird A, Yurgelun-Todd D. 2000. Biological aspects of the link between smoking and depression. *Harv. Rev. Psychiatry* 8:99–110
27. Jorenby DE, Leischow SJ, Nides MA, Rennard SI, Johnston JA, et al. 1999. A controlled trial of sustained-release bupropion, a nicotine patch, or both for smoking cessation. *N. Engl. J. Med.* 340:685–91
28. Durcan MJ, Deener G, White J, Johnston JA, Gonzales D, et al. 2002. The effect of bupropion sustained-release on cigarette craving after smoking cessation. *Clin. Ther.* 24:540–51
29. Shiffman S, Johnston JA, Khayrallah M, Elash CA, Gwaltney CJ, et al. 2000. The effect of bupropion on nicotine craving and withdrawal. *Psychopharmacology* 148:33–40
30. Tashkin D, Kanner R, Bailey W, Buist S, Anderson P, et al. 2001. Smoking cessation in patients with chronic obstructive pulmonary disease: a double-blind, placebo-controlled, randomised trial. *Lancet* 357:1571–75
31. Learned-Coughlin SM, Bergstrom M, Savitcheva I, Ascher J, Schmith VD, Langstrom B. 2003. In vivo activity of bupropion at the human dopamine transporter as measured by positron emission tomography. *Biol. Psychiatry* 54:800–5
32. Argyelan M, Szabo Z, Kanyo B, Tanacs A, Kovacs Z, et al. 2005. Dopamine transporter availability in medication free and in bupropion treated depression: a 99mTc-TRODAT-1 SPECT study. *J. Affect Disord.* 89:115–23

33. Cooper BR, Wang CM, Cox RF, Norton R, Shea V, Ferris RM. 1994. Evidence that the acute behavioral and electrophysiological effects of bupropion (Wellbutrin) are mediated by a noradrenergic mechanism. *Neuropsychopharmacology* 11:133–41
34. Vorel SR, Liu X, Hayes RJ, Spector JA, Gardner EL. 2001. Relapse to cocaine-seeking after hippocampal theta burst stimulation. *Science* 292:1175–78
35. Fryer JD, Lukas RJ. 1999. Noncompetitive functional inhibition at diverse, human nicotinic acetylcholine receptor subtypes by bupropion, phencyclidine, and ibogaine. *J. Pharmacol. Exp. Ther.* 288:88–92
36. Alkondon M, Albuquerque EX. 2005. Nicotinic receptor subtypes in rat hippocampal slices are differentially sensitive to desensitization and early in vivo functional up-regulation by nicotine and to block by bupropion. *J. Pharmacol. Exp. Ther.* 313:740–50
37. Slemmer JE, Martin BR, Damaj MI. 2000. Bupropion is a nicotinic antagonist. *J. Pharmacol. Exp. Ther.* 295:321–27
38. Damaj MI, Carroll FI, Eaton JB, Navarro HA, Blough BE, et al. 2004. Enantioselective effects of hydroxy metabolites of bupropion on behavior and on function of monoamine transporters and nicotinic receptors. *Mol. Pharmacol.* 66:675–82
39. Bondarev ML, Bondareva TS, Young R, Glennon RA. 2003. Behavioral and biochemical investigations of bupropion metabolites. *Eur. J. Pharmacol.* 474:85–93
40. Ahluwalia JS, Harris KJ, Catley D, Okuyemi KS, Mayo MS. 2002. Sustained-release bupropion for smoking cessation in African Americans: a randomized controlled trial. *JAMA* 288:468–74
41. Aubin HJ, Lebargy F, Berlin I, Bidaut-Mazel C, Chemali-Hudry J, Lagrue G. 2004. Efficacy of bupropion and predictors of successful outcome in a sample of French smokers: a randomized placebo-controlled trial. *Addiction* 99:1206–18
42. Gold PB, Rubey RN, Harvey RT. 2002. Naturalistic, self-assignment comparative trial of bupropion SR, a nicotine patch, or both for smoking cessation treatment in primary care. *Am. J. Addict.* 11:315–31
43. Hays JT, Hurt RD, Rigotti NA, Niaura R, Gonzales D, et al. 2001. Sustained-release bupropion for pharmacologic relapse prevention after smoking cessation. a randomized, controlled trial. *Ann. Intern. Med.* 135:423–33
44. Hurt RD, Sachs DP, Glover ED, Offord KP, Johnston JA, et al. 1997. A comparison of sustained-release bupropion and placebo for smoking cessation. *N. Engl. J. Med.* 337:1195–202
45. Jamerson BD, Nides M, Jorenby DE, Donahue R, Garrett P, et al. 2001. Late-term smoking cessation despite initial failure: an evaluation of bupropion sustained release, nicotine patch, combination therapy, and placebo. *Clin. Ther.* 23:744–52
46. Swan GE, McAfee T, Curry SJ, Jack LM, Javitz H, et al. 2003. Effectiveness of bupropion sustained release for smoking cessation in a health care setting: a randomized trial. *Arch. Intern. Med.* 163:2337–44

47. Tonnesen P, Tonstad S, Hjalmarson A, Leborgy F, Van Spiegel PI, et al. 2003. A multicentre, randomized, double-blind, placebo-controlled, 1-year study of bupropion SR for smoking cessation. *J. Intern. Med.* 254:184–92
48. Hughes J, Stead L, Lancaster T. 2004. Antidepressants for smoking cessation. *Cochrane Database Syst. Rev* 4:CD000031
49. Killen JD, Robinson TN, Ammerman S, Hayward C, Rogers J, et al. 2004. Randomized clinical trial of the efficacy of bupropion combined with nicotine patch in the treatment of adolescent smokers. *J. Consult. Clin. Psychol.* 72:729–35
50. Simon JA, Duncan C, Carmody TP, Hudes ES. 2004. Bupropion for smoking cessation: a randomized trial. *Arch. Intern. Med.* 164:1797–803
51. Boshier A, Wilton LV, Shakir SA. 2003. Evaluation of the safety of bupropion (Zyban) for smoking cessation from experience gained in general practice use in England in 2000. *Eur. J. Clin. Pharmacol.* 59:767–73
52. Coe JW, Brooks PR, Vetelino MG, Wirtz MC, Arnold EP, et al. 2005. Varenicline: an $\alpha 4\beta 2$ nicotinic receptor partial agonist for smoking cessation. *J. Med. Chem.* 48:3474–77
53. Papke RL, Heinemann SF. 1994. Partial agonist properties of cytisine on neuronal nicotinic receptors containing the beta 2 subunit. *Mol. Pharmacol.* 45:142–49
54. Bennodorf S, Kempe G, Scharfenberg G, Wendekamm R, Winkelvoss E. 1968. Results of the smoking-habit breaking using cytisin (Tabex). I. *Dtsch. Gesundheitsw.* 23:2092–96
55. Paun D, Franze J. 1968. Breaking the smoking habit using cytisin containing “Tabex” tablets. *Dtsch. Gesundheitsw.* 23:2088–91
56. McCallum SE, Collins AC, Paylor R, Marks MJ. 2006. Deletion of the beta 2 nicotinic acetylcholine receptor subunit alters development of tolerance to nicotine and eliminates receptor upregulation. *Psychopharmacology* 184:314–27
57. Tapper AR, McKinney SL, Nashmi R, Schwarz J, Deshpande P, et al. 2004. Nicotine activation of $\alpha 4^*$ receptors: sufficient for reward, tolerance, and sensitization. *Science* 306:1029–32
58. Walters CL, Cleck JN, Kuo YC, Blendy JA. 2005. Mu-opioid receptor and CREB activation are required for nicotine reward. *Neuron* 46:933–43
59. Oncken C, Watsky E, Reeves K, Anziano R. 2005. *Smoking cessation with varenicline, a selective nicotinic receptor partial agonist: results from a phase 2 study*. Presented at Proc. 11 Annu. Meet. Soc. Res. Nicotine Tobacco, Prague, Czech Rep.
60. Oncken C, Watsky E, Reeves K, Anziano R. 2005. *Varenicline is efficacious and well tolerated in promoting smoking cessation: results from a 7-week, randomized, placebo- and bupropion-controlled trial*. Presented at Proc. 11 Annu. Meet. Soc. Res. Nicotine Tobacco, Prague, Czech Rep.
61. Tonstad S, Hays JT, Jorenby DE, Reeves K, Billing CB, et al. 2005. *Smoking cessation efficacy and safety of an $\alpha 4\beta 2$ nicotinic receptor partial agonist - results from varenicline in cessation therapy: optimizing results*. Presented at Annu. Meet. Am. Heart Assoc., Chicago, IL

62. Cappelleri J, Baker CL, Bushmakin AG, Reeves KR. 2006. *Effect of varenicline on reinforcing effects associated with smoking*. Presented at Proc. 12 Annu. Meet. Soc. Res. Nicotine Tobacco, Orlando, FL
63. Reeves K, Watsky E, Williams K, Azoulay S, Billing B, Gong J. 2006. *The safety of varenicline taken for 52 weeks for smoking cessation*. Presented at Proc. 12 Annu. Meet. Soc. Res. Nicotine Tobacco, Orlando, FL
64. Hieda Y, Keyler DE, Ennifar S, Fattom A, Pentel PR. 2000. Vaccination against nicotine during continued nicotine administration in rats: immunogenicity of the vaccine and effects on nicotine distribution to brain. *Int. J. Immunopharmacol.* 22:809–19
65. Pentel PR, Malin DH, Ennifar S, Hieda Y, Keyler DE, et al. 2000. A nicotine conjugate vaccine reduces nicotine distribution to brain and attenuates its behavioral and cardiovascular effects in rats. *Pharmacol. Biochem. Behav.* 65:191–98
66. Cerny T. 2005. Anti-nicotine vaccination: where are we? *Recent Results Cancer Res.* 166:167–75
67. Le Houezec J. 2005. Why a nicotine vaccine? *Clin. Pharmacol. Ther.* 78:453–55
68. LeSage MG, Keyler DE, Pentel PR. 2006. Current status of immunologic approaches to treating tobacco dependence: vaccines and nicotine-specific antibodies. *Appl. J.* 8:E65–75
69. Lindblom N, de Villiers SH, Kalayanov G, Gordon S, Johansson AM, Svensson TH. 2002. Active immunization against nicotine prevents reinstatement of nicotine-seeking behavior in rats. *Respiration* 69:254–60
70. Lesage MG, Keyler DE, Hieda Y, Collins G, Burroughs D, et al. 2006. Effects of a nicotine conjugate vaccine on the acquisition and maintenance of nicotine self-administration in rats. *Psychopharmacology* 184:409–16
71. Hatsukami DK, Rennard S, Jorenby D, Fiore M, Koopmeiners J, et al. 2005. Safety and immunogenicity of a nicotine conjugate vaccine in current smokers. *Clin. Pharmacol. Ther.* 78:456–67
72. Lindblom N, de Villiers SH, Semenova S, Kalayanov G, Gordon S, et al. 2005. Active immunization against nicotine blocks the reward facilitating effects of nicotine and partially prevents nicotine withdrawal in the rat as measured by dopamine output in the nucleus accumbens, brain reward thresholds and somatic signs. *Naunyn Schmiedeberg's Arch. Pharmacol.* 372:182–94
73. Maurer P, Jennings GT, Willers J, Rohner F, Lindman Y, et al. 2005. A therapeutic vaccine for nicotine dependence: preclinical efficacy, and Phase I safety and immunogenicity. *Eur. J. Immunol.* 35:2031–40
74. Pentel PR, Dufek MB, Roiko SA, LeSage MG, Ross CA, Keyler DE. 2006. *Passive immunization with nicotine-specific monoclonal antibodies as an alternative to vaccination; pharmacokinetic evaluation in rats*. Presented at Proc. 12 Annu. Meet. Soc. Res. Nicotine Tobacco, Orlando, FL
75. Malin DH, Lake JR, Lin A, Saldana M, Balch L, et al. 2001. Passive immunization against nicotine prevents nicotine alleviation of nicotine abstinence syndrome. *Pharmacol. Biochem. Behav.* 68:87–92
76. Malin DH, Newton MA, Goyarzu P, Vega SO, Torok NK, Pentel PR. 2006. *Nicotine place preference conditioned in adolescent rats persists into adulthood and is*

- preventable by immunization against nicotine*. Presented at Proc. 12 Annu. Meet. Soc. Res. Nicotine Tobacco, Orlando, FL
77. Chen JP, Paredes W, Li J, Smith D, Lowinson J, Gardner EL. 1990. Delta 9-tetrahydrocannabinol produces naloxone-blockable enhancement of presynaptic basal dopamine efflux in nucleus accumbens of conscious, freely-moving rats as measured by intracerebral microdialysis. *Psychopharmacology* 102:156–62
 78. Tanda G, Pontieri FE, Di Chiara G. 1997. Cannabinoid and heroin activation of mesolimbic dopamine transmission by a common mu1 opioid receptor mechanism. *Science* 276:2048–50
 79. French ED. 1997. delta9-Tetrahydrocannabinol excites rat VTA dopamine neurons through activation of cannabinoid CB1 but not opioid receptors. *Neurosci. Lett.* 226:159–62
 80. Gonzalez S, Cascio MG, Fernandez-Ruiz J, Fezza F, Di Marzo V, Ramos JA. 2002. Changes in endocannabinoid contents in the brain of rats chronically exposed to nicotine, ethanol or cocaine. *Brain Res.* 954:73–81
 81. Shoemaker JL, Ruckle MB, Mayeux PR, Prather PL. 2005. Agonist-directed trafficking of response by endocannabinoids acting at CB2 receptors. *J. Pharmacol. Exp. Ther.* 315:828–38
 82. Steffens M, Zentner J, Honegger J, Feuerstein TJ. 2005. Binding affinity and agonist activity of putative endogenous cannabinoids at the human neocortical CB1 receptor. *Biochem. Pharmacol.* 69:169–78
 83. Cohen C, Perrault G, Voltz C, Steinberg R, Soubrie P. 2002. SR141716, a central cannabinoid (CB1) receptor antagonist, blocks the motivational and dopamine-releasing effects of nicotine in rats. *Behav. Pharmacol.* 13:451–63
 84. Cohen C, Perrault G, Griebel G, Soubrie P. 2005. Nicotine-associated cues maintain nicotine-seeking behavior in rats several weeks after nicotine withdrawal: reversal by the cannabinoid (CB1) receptor antagonist, rimonabant (SR141716). *Neuropsychopharmacology* 30:145–55
 85. Le Foll B, Goldberg SR. 2004. Rimonabant, a CB1 antagonist, blocks nicotine-conditioned place preferences. *Neuroreport* 15:2139–43
 86. Castane A, Valjent E, Ledent C, Parmentier M, Maldonado R, Valverde O. 2002. Lack of CB1 cannabinoid receptors modifies nicotine behavioural responses, but not nicotine abstinence. *Neuropharmacology* 43:857–67
 87. Cossu G, Ledent C, Fattore L, Imperato A, Bohme GA, et al. 2001. Cannabinoid CB1 receptor knockout mice fail to self-administer morphine but not other drugs of abuse. *Behav. Brain Res.* 118:61–65
 88. Anthenelli RM, Despres JP. 2004. *Effects of rimonabant in the reduction of major cardiovascular risk factors. Results from the STRATUS-US trial (smoking cessation in smokers motivated to quit)*. Presented at Am. College Cardiol. 53rd Annu. Sci. Session, Rapid News Summaries, New Orleans, LA
 89. Anthenelli RM. 2005. *Rimonabant in smoking abstinence: the STRATUS-program*. Presented at Proc. 11 Annu. Meet. Soc. Res. Nicotine Tobacco, Prague, Czech Rep.
 90. Hall SM, Reus VI, Munoz RF, Sees KL, Humfleet G, et al. 1998. Nortriptyline and cognitive-behavioral therapy in the treatment of cigarette smoking. *Arch. Gen. Psychiatry* 55:683–90

91. Prochazka AV, Weaver MJ, Keller RT, Fryer GE, Licari PA, Lofaso D. 1998. A randomized trial of nortriptyline for smoking cessation. *Arch. Intern. Med.* 158:2035–39
92. da Costa CL, Younes RN, Lourenco MT. 2002. Stopping smoking: a prospective, randomized, double-blind study comparing nortriptyline to placebo. *Chest* 122:403–8
93. Hall SM, Humfleet GL, Reus VI, Munoz RF, Hartz DT, Maude-Griffin R. 2002. Psychological intervention and antidepressant treatment in smoking cessation. *Arch. Gen. Psychiatry* 59:930–36
94. Hughes JR, Stead LF, Lancaster T. 2005. Nortriptyline for smoking cessation: a review. *Nicotine Tob. Res.* 7:491–99
95. Gossop M. 1988. Clonidine and the treatment of the opiate withdrawal syndrome. *Drug Alcohol. Depend.* 21:253–59
96. Manhem P, Nilsson LH, Moberg AL, Wadstein J, Hokfelt B. 1985. Alcohol withdrawal: effects of clonidine treatment on sympathetic activity, the renin-aldosterone system, and clinical symptoms. *Alcohol. Clin. Exp. Res.* 9:238–43
97. Glassman AH, Jackson WK, Walsh BT, Roose SP, Rosenfeld B. 1984. Cigarette craving, smoking withdrawal, and clonidine. *Science* 226:864–66
98. Ornish SA, Zisook S, McAdams LA. 1988. Effects of transdermal clonidine treatment on withdrawal symptoms associated with smoking cessation. A randomized, controlled trial. *Arch. Intern. Med.* 148:2027–31
99. Gourlay SG, Stead LF, Benowitz NL. 2004. Clonidine for smoking cessation. *Cochrane Database Syst. Rev.* 2:CD000058
100. Shytle RD, Penny E, Silver AA, Goldman J, Sanberg PR. 2002. Mecamylamine (Inversine): an old antihypertensive with new research directions. *J. Hum. Hypertens* 16:453–57
101. Rose JE, Behm FM, Westman EC, Levin ED, Stein RM, Ripka GV. 1994. Mecamylamine combined with nicotine skin patch facilitates smoking cessation beyond nicotine patch treatment alone. *Clin. Pharmacol. Ther.* 56:86–99
102. Rose JE, Behm FM, Westman EC. 1998. Nicotine-mecamylamine treatment for smoking cessation: the role of precessation therapy. *Exp. Clin. Psychopharmacol.* 6:331–43
103. Eissenberg T, Griffiths RR, Stitzer ML. 1996. Mecamylamine does not precipitate withdrawal in cigarette smokers. *Psychopharmacology* 127:328–36
104. Fowler JS, Logan J, Wang GJ, Volkow ND. 2003. Monoamine oxidase and cigarette smoking. *Neurotoxicology* 24:75–82
105. Berlin I, Said S, Spreux-Varoquaux O, Launay JM, Olivares R, et al. 1995. A reversible monoamine oxidase A inhibitor (moclobemide) facilitates smoking cessation and abstinence in heavy, dependent smokers. *Clin. Pharmacol. Ther.* 58:444–52
106. George TP, Vessicchio JC, Termine A, Jatlow PI, Kosten TR, O'Malley SS. 2003. A preliminary placebo-controlled trial of selegiline hydrochloride for smoking cessation. *Biol. Psychiatry* 53:136–43
107. Berlin I, Aubin HJ, Pedarriosse AM, Rames A, Lancrenon S, Lagrue G. 2002. Lazabemide, a selective, reversible monoamine oxidase B inhibitor, as an aid to smoking cessation. *Addiction* 97:1347–54

108. Andreoli M, Tessari M, Pilla M, Valerio E, Hagan JJ, Heidbreder CA. 2003. Selective antagonism at dopamine D3 receptors prevents nicotine-triggered relapse to nicotine-seeking behavior. *Neuropsychopharmacology* 28:1272–80
109. Barrett AC, Miller JR, Dohrmann JM, Caine SB. 2004. Effects of dopamine indirect agonists and selective D1-like and D2-like agonists and antagonists on cocaine self-administration and food maintained responding in rats. *Neuropharmacology* 47(Suppl. 1):256–73
110. Di Ciano P, Underwood RJ, Hagan JJ, Everitt BJ. 2003. Attenuation of cue-controlled cocaine-seeking by a selective D3 dopamine receptor antagonist SB-277011-A. *Neuropsychopharmacology* 28:329–38
111. Le Foll B, Sokoloff P, Stark H, Goldberg SR. 2005. Dopamine D3 receptor ligands block nicotine-induced conditioned place preferences through a mechanism that does not involve discriminative-stimulus or antidepressant-like effects. *Neuropsychopharmacology* 30:720–30
112. Pilla M, Perachon S, Sautel F, Garrido F, Mann A, et al. 1999. Selective inhibition of cocaine-seeking behavior by a partial dopamine D3 receptor agonist. *Nature* 400:371–75
113. Duarte C, Lefebvre C, Chaperon F, Hamon M, Thiebot MH. 2003. Effects of a dopamine D3 receptor ligand, BP 897, on acquisition and expression of food-, morphine-, and cocaine-induced conditioned place preference, and food-seeking behavior in rats. *Neuropsychopharmacology* 28:1903–15
114. Le Foll B, Goldberg SR, Sokoloff P. 2005. The dopamine D3 receptor and drug dependence: effects on reward or beyond? *Neuropharmacology* 49:525–41
115. Benowitz NL, Jacob P. 1994. Metabolism of nicotine to cotinine studied by a dual stable isotope method. *Clin. Pharmacol. Ther.* 56:483–93
116. Messina ES, Tyndale RF, Sellers EM. 1997. A major role for CYP2A6 in nicotine C-oxidation by human liver microsomes. *J. Pharmacol. Exp. Ther.* 282:1608–14
117. Brandange S, Lindblom L. 1979. The enzyme “aldehyde oxidase” is an iminium oxidase. Reaction with nicotine delta 1'(5') iminium ion. *Biochem. Biophys. Res. Commun.* 91:991–96
118. Murphy S, Raulinaitis V, Brown KM. 2005. Nicotine 5'-oxidation and methyl oxidation by P450 2A enzymes. *Drug Metab. Dispos.* 33(8):1166–73
119. Williams DE, Shigenaga MK, Castagnoli NJ. 1990. The role of cytochromes P-450 and flavin-containing monooxygenase in the metabolism of (S)-nicotine by rabbit lung. *Drug Metab. Dispos.* 18:418–28
120. Yamazaki H, Inoue K, Hashimoto M, Shimada T. 1999. Roles of CYP2A6 and CYP2B6 in nicotine C-oxidation by human liver microsomes. *Arch. Toxicol.* 73:65–70
121. Nakajima M, Yamamoto T, Nunoya K, Yokoi T, Nagashima K, et al. 1996. Role of human cytochrome P4502A6 in C-oxidation of nicotine. *Drug Metab. Dispos.* 24:1212–17
122. Cushman JR, Park SB, Yang ZC, Wrighton SA, Jacob P, Benowitz NL. 1992. Metabolism of nicotine by human liver microsomes: stereoselective formation of trans-nicotine N'-oxide. *Chem. Res. Toxicol.* 5:639–46

123. Park SB, Jacob P, Benowitz NL, Cashman JR. 1993. Stereoselective metabolism of (S)-(-)-nicotine in humans: formation of trans-(S)-(-)-nicotine N-1'-oxide. *Chem. Res. Toxicol.* 6:880-88
124. Kuehl GE, Murphy SE. 2003. N-glucuronidation of nicotine and cotinine by human liver microsomes and heterologously expressed UDP-glucuronosyltransferases. *Drug Metab. Dispos.* 31:1361-68
125. Nakajima M, Yamamoto T, Nunoya K, Yokoi T, Nagashima K, et al. 1996. Characterization of CYP2A6 involved in 3'-hydroxylation of cotinine in human liver microsomes. *J. Pharmacol. Exp. Ther.* 277:1010-15
126. Yamanaka H, Nakajima M, Nishimura K, Yoshida R, Fukami T, et al. 2004. Metabolic profile of nicotine in subjects whose CYP2A6 gene is deleted. *Eur. J. Pharm. Sci.* 22:419-25
127. Lerman C, Tyndale R, Patterson F, Wileyto EP, Shields PG, et al. 2006. Nicotine metabolite ratio predicts the efficacy of transdermal nicotine for smoking cessation. *Clin. Pharmacol. Ther.* 79:600-8
128. Malaiyandi V, Lerman C, Benowitz NL, Jepson C, Patterson F, Tyndale RF. 2006. Impact of CYP2A6 genotype on pretreatment smoking behavior and nicotine levels from and usage of nicotine replacement therapy. *Mol. Psychiatry* 11(4):400-9
129. Nakajima M, Kwon JT, Tanaka N, Zenta T, Yamamoto Y, et al. 2001. Relationship between interindividual differences in nicotine metabolism and CYP2A6 genetic polymorphism in humans. *Clin. Pharmacol. Ther.* 69:72-78
130. Oscarson M, McLellan RA, Gullsten H, Yue QY, Lang MA, et al. 1999. Characterisation and PCR-based detection of a CYP2A6 gene deletion found at a high frequency in a Chinese population. *FEBS Lett.* 448:105-10
131. Dempsey D, Tutka P, Jacob P, Allen F, Schoedel K, et al. 2004. Nicotine metabolite ratio as an index of cytochrome P450 2A6 metabolic activity. *Clin. Pharmacol. Ther.* 76:64-72
132. Rao Y, Hoffmann E, Zia M, Bodin L, Zeman M, et al. 2000. Duplications and defects in the CYP2A6 gene: identification, genotyping, and in vivo effects on smoking. *Mol. Pharmacol.* 58:747-55
133. Schoedel KA, Hoffmann EB, Rao Y, Sellers EM, Tyndale RF. 2004. Ethnic variation in CYP2A6 and association of genetically slow nicotine metabolism and smoking in adult Caucasians. *Pharmacogenetics* 14:615-26
134. Gu DF, Hinks LJ, Morton NE, Day IN. 2000. The use of long PCR to confirm three common alleles at the CYP2A6 locus and the relationship between genotype and smoking habit. *Ann. Hum. Genet.* 64:383-90
135. Draper AJ, Madan A, Parkinson A. 1997. Inhibition of coumarin 7-hydroxylase activity in human liver microsomes. *Arch. Biochem. Biophys.* 341:47-61
136. Koenigs LL, Peter RM, Thompson SJ, Rettie AE, Trager WF. 1997. Mechanism-based inactivation of human liver cytochrome P450 2A6 by 8-methoxypsoralen. *Drug Metab. Dispos.* 25:1407-15
137. Maenpaa J, Juvonen R, Raunio H, Rautio A, Pelkonen O. 1994. Metabolic interactions of methoxsalen and coumarin in humans and mice. *Biochem. Pharmacol.* 48:1363-69

138. Zhang W, Kilicarslan T, Tyndale RF, Sellers EM. 2001. Evaluation of methoxsalen, tranlylcypromine, and tryptamine as specific and selective CYP2A6 inhibitors in vitro. *Drug Metab. Dispos.* 29:897-902
139. Tassaneeyakul W, Birkett DJ, Veronese ME, McManus ME, Tukey RH, et al. 1993. Specificity of substrate and inhibitor probes for human cytochromes P450 1A1 and 1A2. *J. Pharmacol. Exp. Ther.* 265:401-7
140. Zhang W, Kilicarslan T, Tyndale RF, Seller EM. 2000. Identification of selective and specific inhibitors of CYP2A6. *Clin. Pharmacol. Ther.* 67:166 (Abstr.)
141. Taavitsainen P, Juvonen R, Pelkonen O. 2001. In vitro inhibition of cytochrome P450 enzymes in human liver microsomes by a potent CYP2A6 inhibitor, trans-2-phenylcyclopropylamine (tranlylcypromine), and its nonamine analog, cyclopropylbenzene. *Drug Metab. Dispos.* 29:217-22
142. Benowitz NL, Jacob P, Savanapridi C. 1987. Determinants of nicotine intake while chewing nicotine polacrilex gum. *Clin. Pharmacol. Ther.* 41:467-73
143. Benowitz NL, Jacob P, Denaro C, Jenkins R. 1991. Stable isotope studies of nicotine kinetics and bioavailability. *Clin. Pharmacol. Ther.* 49:270-77
144. Sellers EM, Dortok D, Tyndale RF. 2002. CYP2A6 inhibition increases plasma nicotine concentrations during nicotine patch and gum. *Clin. Pharmacol. Ther.* 71:17
145. Tyndale RF, Li Y, Kaplan HL, Seller EM. 1999. Inhibition of nicotine's metabolism: a potential new treatment for tobacco dependence. *Clin. Pharmacol. Ther.* 65:145 (Abstr.)
146. Sellers EM, Kaplan HL, Tyndale RF. 2000. Inhibition of cytochrome P450 2A6 increases nicotine's oral bioavailability and decreases smoking. *Clin. Pharmacol. Ther.* 68:35-43
147. Maser E, Richter E, Friebertshauser J. 1996. The identification of 11 beta-hydroxysteroid dehydrogenase as carbonyl reductase of the tobacco-specific nitrosamine 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone. *Eur. J. Biochem.* 238:484-89
148. Hecht SS, Trushin N, Reid-Quinn CA, Burak ES, Jones AB, et al. 1993. Metabolism of the tobacco-specific nitrosamine 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone in the patas monkey: pharmacokinetics and characterization of glucuronide metabolites. *Carcinogenesis* 14:229-36
149. Hecht SS, Spratt TE, Trushin N. 1988. Evidence for 4-(3-pyridyl)-4-oxobutylolation of DNA in F344 rats treated with the tobacco-specific nitrosamines 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone and N'-nitrosonornicotine. *Carcinogenesis* 9:161-65
150. Peterson LA, Carmella SG, Hecht SS. 1990. Investigations of metabolic precursors to hemoglobin and DNA adducts of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone. *Carcinogenesis* 11:1329-33
151. Nishikawa A, Mori Y, Lee IS, Tanaka T, Hirose M. 2004. Cigarette smoking, metabolic activation and carcinogenesis. *Curr. Drug Metab.* 5:363-73
152. Fujieda M, Yamazaki H, Saito T, Kiyotani K, Gyamfi MA, et al. 2004. Evaluation of CYP2A6 genetic polymorphisms as determinants of smoking behavior and tobacco-related lung cancer risk in male Japanese smokers. *Carcinogenesis* 25:2451-58

153. Hecht SS, Chen CB, Hirota N, Orna RM, Tso TC, Hoffmann D. 1978. Tobacco-specific nitrosamines: formation from nicotine in vitro and during tobacco curing and carcinogenicity in strain A mice. *J Natl. Cancer Inst.* 60:819–24
154. Hecht SS, Isaacs S, Trushin N. 1994. Lung tumor induction in A/J mice by the tobacco smoke carcinogens 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone and benzo[a]pyrene: a potentially useful model for evaluation of chemopreventive agents. *Carcinogenesis* 15:2721–25
155. Okubo C, Morishita Y, Minami Y, Ishiyama T, Kano J, et al. 2005. Phenotypic characteristics of mouse lung adenoma induced by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone. *Mol Carcinog* 42:121–26
156. Miyazaki M, Yamazaki H, Takeuchi H, Saoo K, Yokohira M, et al. 2005. Mechanisms of chemopreventive effects of 8-methoxypsoralen against 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone-induced mouse lung adenomas. *Carcinogenesis* 26:1947–55
157. Takeuchi H, Saoo K, Yokohira M, Ikeda M, Maeta H, et al. 2003. Pretreatment with 8-methoxypsoralen, a potent human CYP2A6 inhibitor, strongly inhibits lung tumorigenesis induced by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone in female A/J mice. *Cancer Res.* 63:7581–83
158. Sellers EM, Ramamoorthy Y, Zeman MV, Djordjevic MV, Tyndale RF. 2003. The effect of methoxsalen on nicotine and 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) metabolism in vivo. *Nicotine Tob. Res.* 5:891–99
159. Poso A, Gynther J, Juvonen R. 2001. A comparative molecular field analysis of cytochrome P450 2A5 and 2A6 inhibitors. *J. Comput. Aided. Mol. Des.* 15:195–202
160. Rahnasto M, Raunio H, Poso A, Wittekindt C, Juvonen RO. 2005. Quantitative structure-activity relationship analysis of inhibitors of the nicotine metabolizing CYP2A6 enzyme. *J. Med. Chem.* 48:440–49
161. Yano JK, Hsu MH, Griffin KJ, Stout CD, Johnson EF. 2005. Structures of human microsomal cytochrome P450 2A6 complexed with coumarin and methoxsalen. *Nat. Struct. Mol. Biol.* 12:822–23
162. Asikainen A, Tarhanen J, Poso A, Pasanen M, Alhava E, Juvonen RO. 2003. Predictive value of comparative molecular field analysis modelling of naphthalene inhibition of human CYP2A6 and mouse CYP2A5 enzymes. *Toxicol. In Vitro* 17:449–55
163. Denton TT, Zhang X, Cashman JR. 2004. Nicotine-related alkaloids and metabolites as inhibitors of human cytochrome P-450 2A6. *Biochem. Pharmacol.* 67:751–56
164. Fujita K, Kamataki T. 2001. Screening of organosulfur compounds as inhibitors of human CYP2A6. *Drug Metab. Dispos.* 29:983–89
165. Denton TT, Zhang X, Cashman JR. 2005. 5-substituted, 6-substituted, and unsubstituted 3-heteroaromatic pyridine analogues of nicotine as selective inhibitors of cytochrome P-450 2A6. *J. Med. Chem.* 48:224–39
166. Nakayama H, Okuda H, Nakashima T, Imaoka S, Funae Y. 1993. Nicotine metabolism by rat hepatic cytochrome P450s. *Biochem. Pharmacol.* 45:2554–56

167. Schoedel KA, Tyndale RF. 2003. Induction of nicotine-metabolizing CYP2B1 by ethanol and ethanol-metabolizing CYP2E1 by nicotine: summary and implications. *Biochim. Biophys. Acta* 1619:283–90
168. Katoh M, Matsui T, Nakajima M, Tateno C, Kataoka M, et al. 2004. Expression of human cytochromes P450 in chimeric mice with humanized liver. *Drug Metab. Dispos.* 32:1402–10
169. Katoh M, Matsui T, Okumura H, Nakajima M, Nishimura M, et al. 2005. Expression of human phase II enzymes in chimeric mice with humanized liver. *Drug Metab. Dispos.* 33:1333–40
170. Nishimura M, Yoshitsugu H, Yokoi T, Tateno C, Kataoka M, et al. 2005. Evaluation of mRNA expression of human drug-metabolizing enzymes and transporters in chimeric mouse with humanized liver. *Xenobiotica* 35:877–90
171. Tateno C, Yoshizane Y, Saito N, Kataoka M, Utoh R, et al. 2004. Near completely humanized liver in mice shows human-type metabolic responses to drugs. *Am. J. Pathol.* 165:901–12
172. Rose JE. 2006. Nicotine and nonnicotine factors in cigarette addiction. *Psychopharmacology* 184(3–4):274–85
173. Rose JE, Behm FM, Westman EC, Bates JE, Salley A. 2003. Pharmacologic and sensorimotor components of satiation in cigarette smoking. *Pharmacol. Biochem. Behav.* 76:243–50
174. Rose JE, Behm FM, Westman EC, Johnson M. 2000. Dissociating nicotine and nonnicotine components of cigarette smoking. *Pharmacol. Biochem. Behav.* 67:71–81
175. Rose JE, Tashkin DP, Ertle A, Zinser MC, Lafer R. 1985. Sensory blockade of smoking satisfaction. *Pharmacol. Biochem. Behav.* 23:289–93
176. Rose JE, Zinser MC, Tashkin DP, Newcomb R, Ertle A. 1984. Subjective response to cigarette smoking following airway anesthetization. *Addict. Behav.* 9:211–15
177. Perkins KA, Gerlach D, Vender J, Grobe J, Meeker J, Hutchison S. 2001. Sex differences in the subjective and reinforcing effects of visual and olfactory cigarette smoke stimuli. *Nicotine Tob. Res.* 3:141–50
178. Balfour DJ, Fagerstrom KO. 1996. Pharmacology of nicotine and its therapeutic use in smoking cessation and neurodegenerative disorders. *Pharmacol. Ther.* 72:51–81
179. Behm FM, Levin ED, Lee YK, Rose JE. 1990. Low-nicotine regenerated smoke aerosol reduces desire for cigarettes. *J. Subst. Abuse* 2:237–47
180. Behm FM, Schur C, Levin ED, Tashkin DP, Rose JE. 1993. Clinical evaluation of a citric acid inhaler for smoking cessation. *Drug Alcohol. Depend.* 31:131–38
181. Lancaster T, Stead LF. 2005. Individual behavioural counseling for smoking cessation. *Cochrane Database Syst. Rev.* 2:CD001292
182. Stead LF, Lancaster T. 2005. Group behavior therapy programmes for smoking cessation. *Cochrane Database Syst. Rev.* 2:CD001007
183. Hajek P, Stead LF, West R, Jarvis M. 2005. Relapse prevention interventions for smoking cessation. *Cochrane Database Syst. Rev.* 1:CD003999



Contents

Allosteric Modulation of G Protein–Coupled Receptors <i>Lauren T. May, Katie Leach, Patrick M. Sexton, and Arthur Christopoulos</i>	1
Pharmacogenomic and Structural Analysis of Constitutive G Protein–Coupled Receptor Activity <i>Martine J. Smit, Henry F. Vischer, Remko A. Bakker, Aldo Jongejan, Henk Timmerman, Leonardo Pardo, and Rob Leurs</i>	53
Cell Survival Responses to Environmental Stresses Via the Keap1-Nrf2-ARE Pathway <i>Thomas W. Kensler, Nobunao Wakabayashi, and Shyam Biswal</i>	89
Cell Signaling and Neuronal Death <i>Makoto R. Hara and Solomon H. Snyder</i>	117
Mitochondrial Oxidative Stress: Implications for Cell Death <i>Sten Orrenius, Vladimir Gogvadze, and Boris Zhivotovsky</i>	143
AMP-Activated Protein Kinase as a Drug Target <i>D. Grabame Hardie</i>	185
Intracellular Targets of Matrix Metalloproteinase-2 in Cardiac Disease: Rationale and Therapeutic Approaches <i>Richard Schulz</i>	211
Arsenic: Signal Transduction, Transcription Factor, and Biotransformation Involved in Cellular Response and Toxicity <i>Yoshito Kumagai and Daigo Sumi</i>	243
Aldo-Keto Reductases and Bioactivation/Detoxication <i>Yi Jin and Trevor M. Penning</i>	263
Carbonyl Reductases: The Complex Relationships of Mammalian Carbonyl- and Quinone-Reducing Enzymes and Their Role in Physiology <i>Udo Oppermann</i>	293
Drug Targeting to the Brain <i>A.G. de Boer and P.J. Gaillard</i>	323
Mechanism-Based Pharmacokinetic-Pharmacodynamic Modeling: Biophase Distribution, Receptor Theory, and Dynamical Systems Analysis <i>Meindert Danhof, Joost de Jongh, Elizabeth C.M. De Lange, Oscar Della Pasqua, Bart A. Ploeger, and Rob A. Voskuyl</i>	357

The Functional Impact of SLC6 Transporter Genetic Variation <i>Maureen K. Habn and Randy D. Blakely</i>	401
mTOR Pathway as a Target in Tissue Hypertrophy <i>Chung-Han Lee, Ken Inoki, and Kun-Liang Guan</i>	443
Diseases Caused by Defects in the Visual Cycle: Retinoids as Potential Therapeutic Agents <i>Gabriel H. Travis, Marcin Golczak, Alexander R. Moise, and Krzysztof Palczewski</i> ...	469
Idiosyncratic Drug Reactions: Current Understanding <i>Jack Uetrecht</i>	513
Non-Nicotinic Therapies for Smoking Cessation <i>Eric C.K. Siu and Rachel F. Tyndale</i>	541
The Obesity Epidemic: Current and Future Pharmacological Treatments <i>Karl G. Hofbauer, Janet R. Nicholson, and Olivier Boss</i>	565
Circadian Rhythms: Mechanisms and Therapeutic Implications <i>Francis Levi and Ueli Schibler</i>	593
Targeting Antioxidants to Mitochondria by Conjugation to Lipophilic Cations <i>Michael P. Murphy and Robin A.J. Smith</i>	629
Acute Effects of Estrogen on Neuronal Physiology <i>Catherine S. Woolley</i>	657
New Insights into the Mechanism of Action of Amphetamines <i>Annette E. Fleckenstein, Trent J. Volz, Evan L. Riddle, James W. Gibb, and Glen R. Hanson</i>	681
Nicotinic Acetylcholine Receptors and Nicotinic Cholinergic Mechanisms of the Central Nervous System <i>John A. Dani and Daniel Bertrand</i>	699
Contrasting Actions of Endothelin ET _A and ET _B Receptors in Cardiovascular Disease <i>Markus P. Schneider, Erika I. Boesen, and David M. Pollock</i>	731

Indexes

Cumulative Index of Contributing Authors, Volumes 43–47	761
Cumulative Index of Chapter Titles, Volumes 43–47	764

Errata

An online log of corrections to *Annual Review of Pharmacology and Toxicology* chapters (if any, 1997 to the present) may be found at
<http://pharmtox.annualreviews.org/errata.shtml>